

CURRENT EVIDENCE ON THE BENEFICIAL PROPERTIES OF LACTOBACILLUS SPECIES IN LACTOSE INTOLERANCE

M. Brigida¹, A. Saviano², G. De Carlo³, A. Piccioni⁴, V. Ojetti⁵

¹Unit of Gastroenterology, Department of Systems Medicine, Tor Vergata University, Rome, Italy ²Università Cattolica del Sacro Cuore, Rome, Italy ³Department of Digestive Diseases, Campus Bio Medico University of Rome, Rome, Italy ⁴Fondazione Policlinico Universitario A. Gemelli, IRCCS, Rome, Italy ⁵Department of Internal Medicine, San Carlo di Nancy Hospital, Rome, Italy

Corresponding Author: Mattia Brigida, MD; email: mattia.brigida@ptvonline.it

Abstract – *Objectives:* Lactose intolerance (LI) is a condition characterized by absent or strongly decreased levels of intestinal lactase. This is a key enzyme for the lactose hydrolysis to monosaccharides, glucose, and galactose. Colonic bacteria metabolize the unabsorbed lactose, producing short-chain fatty acids and gas and resulting in nausea, bloating and diarrhea. Probiotics are gaining interest as a potential compensation for lactase insufficiency, particularly the predominant ones in the gastrointestinal microbiota, such as Bifidobacterium and Lactobacillus. Among the latter species, medical attention is nowadays focusing on a gram-positive strain, *Lactobacillus Reuteri*, which is able to stabilize intestinal permeability and to reduce flatulence, diarrhea and nausea. The aim of this review is to collect and summarize the current evidence concerning the beneficial effects of *L. Reuteri* in the treatment of LI.

Materials and Methods: We conducted a thorough search on PubMed concerning evidence spanning from 2011 to 2021.

Results: Lactobacillus Reuteri has been demonstrated to stabilize intestinal permeability and to be effective in attenuating clinical signs and symptoms of LI. In particular, one study shows that Lactobacillus reuteri treatment significantly improves lactose digestion with respect to placebo; however, its effects were lower than those observed with tilactase supplementation.

Conclusions: Although literature is scant on this matter, *Lactobacillus Reuteri* seems to meet the requirements to be safely used in managing symptoms of lactose intolerance.

Keywords: Lactobacillus spp, Lactobacillus Reuteri (DSM 17938), Lactose intolerance, Probiotics, Microbiota, Lactase.

INTRODUCTION

Lactose intolerance (LI) is a lifelong disorder due to a reduced amount and/or a decreased activity of the enzyme lactase located in the brush border (microvilli) of the small intestine enterocytes, which causes a malabsorption of dietary lactose in the small intestine and results in a subsequent accumulation of undigested lactose in the colon. As a consequence, this causes an increase in fermentation by the resident flora and exerts locally osmotic effect that leads to gastrointestinal symptoms, such as bloating, excessive flatus, abdominal pain and diarrhea.

COSO This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License

Three different forms of hypolactasia, or lactase deficiency, exist: primary, secondary, and congenital. Congenital lactase deficiency is an extremely rare autosomal recessive disorder associated with a decreased lactase activity¹. It is a lifelong disorder characterized by growth retardation, where the very first exposure to lactose at birth leads to infantile diarrhea. The only way to treat this disease is to completely avoid lactose from birth. Primary lactase deficiency (lactase non-persistence) occurs in the majority of humans: the expression of lactase in the gut, maximum at birth, undergoes a gradual and progressive reduction after weaning in most mammals (up to 75-90% of the enzyme is lost), resulting in malabsorption of the lactose usually occurring not earlier than 6-7 years of age, and sometimes much later², showing an increase in prevalence even in the age groups over 65 years³. Acquired or secondary lactase deficiency represents the loss of lactase activity in a subject with prior normal levels of lactase. It is the consequence of diseases that damage the intestinal epithelium, e.g., untreated coeliac disease or intestinal inflammation^{4,5} or can result from small intestine resections, and from gastrectomy⁶.

Lactose intolerance can be usually diagnosed by conducting a thorough medical history supported by dietary changes. Patients usually have a history of flatulence and/or diarrhea after ingestion of milk and its derivatives. In case of diagnostic suspicion, it is necessary to carry out the hydrogen breath test, 50 grams of lactose are administered orally to the patient; this is followed by the measurement of the hydrogen produced by the bacterial metabolism of undigested lactose using a respirometer at a distance of 2, 3 and 4 hours from ingestion. The affected patients most of the times have an increase in exhaled H2> 20 parts per million from baseline. Sensitivity and specificity are > $95\%^7$.

Carbohydrate malabsorption, in the specific case of lactase deficiency, is easily controlled by following a lactose-free diet. The key treatment of lactose intolerance is to reduce the dietary quota of lactose until it is completely abolished. It is useful to use delactosed milk, containing a reduced percentage of lactose, obtained through enzymatic hydrolysis, to allow the intake of milk nutrients (calcium and glucose and galactose, that are the simple sugars constituents of lactose). Yogurt is usually well tolerated, since it contains a fair amount of intrinsic lactase produced by the strains of *Streptococcus thermophilus* contained therein⁸. As mentioned by Marteau et al⁹, high levels of lactase are contained within yogurt bacteria, and this is quickly released when bile salts lyse bacteria in the gastrointestinal tract. Consequently, tolerance to yogurt shown by lactase deficient people seems to be due to the lactase content of the bacterial strains and to a lesser extent to the action of the bacteria themselves. In addition, other probiotics containing lactase, such as Lactobacillus acidophilus may also be effective, but their lesser efficiency compared to yogurt bacteria can be explained by a higher resistance to bile¹⁰.

In this narrative review, we collected recent updates (Table I) on the beneficial properties of *Lactobacillus spp*, particularly *L. reuteri* 17938, and their potential niche in the management of lactose intolerant patients.

Role of Probiotics in the Management of Lactose Intolerance

Recently, the effectiveness of probiotics in the treatment of LI was examined using 15 randomized double-blind studies evaluating probiotic strains with the greatest number of proven benefits¹¹. Various degrees of efficacy are reported in the results, but an overall positive relationship exists between probiotics and lactose intolerance. In particular, *Bifidobacteroium longum*, a Gram-positive, non-motile, non-sporulating lactic-acid bacteria, modifies the amount and metabolic activities of the colonic microbiota and alleviates symptoms in lactose-intolerant subjects. Supplementation acts by changing colonic microbiota, leading to attenuation of LI symptoms. Probiotic supplementation of *Bifidobacterium animalis*, a Gram-positive, anaerobic, rod-shaped bacteria found in the large intestine, can improve the symptoms of lactose intolerance in adults improving overall gastrointestinal transit time¹². There is limited evidence to suggest that *Lactobacillus bulgaricus*, a Gram-positive, facultatively anaerobic, non-motile lactic acid bacteria, is effective in eliminating LI symptoms by improving lactose digestion. *Lactobacillus acidophilus*, a Gram-positive bacterium, analyzed in nine studies, did not show significant potential to

	TABLE 1. SUMMAR "LACTOBACILLUS R	IV OF THE MOST R	ELEVANT BI 38)", "LACI	BLIOGRAPHY CONSIDERED FOR THIS REVIEW (KEYWORDS USED: OSE INTOLERANCE", "PROBIOTICS", "MICROBIOTA", "LACTASE").
First Author	Type	-•Year of publication	No. of patients	Main Findings
Oak SJ ¹¹	Systematic	2018 review	~	The effectiveness of probiotics in the treatment of lactose intolerance was assessed using 15 randomized double-blind trials. Eight strains of probiotics with the highest number of proven benefits were evaluated. Different degrees of efficacy were shown but an overall positive relationship was found between lactose intolerance and probiotics.
Marteau P ¹²	Double-blind,	2002 cross-over study	36	Bifidobacterium animalis DN-173 010 administration in healthy women reduces the colonic transit time and this fact is not explained by modifications of the secondary bile acids or faecal bacterial mass.
Yesovitch R ¹³	Pilot study	2004	10	Direct consumption of the probiotic VSL3 may not improve parameters of lactose maldigestion without metabolic activation.
Ojetti V ¹⁴	Randomized	2010 controlled trial	60	In lactose intolerants, tilactase strongly improves both gastrointestinal symptoms lactose and breath test results after lactose ingestion compared to placebo. <i>Lactobacillus reuteri</i> also is effective, but lesserthan tilactase. This probiotic may represent an interesting treatment option for lactose intolerants due to its simple use, and its effect may even last in the time after stopping its administration.
Duar RM16	Narrative Review	2017	~	An elucidation on the evolution and natural history of Lactobacillus genus: phylogenetic, genomic and metabolic metadata
Mu Q21	Narrative Review	2018	~	A detailed explanation of the different ways through which inflammatory diseases, including those located in the gut, may be ameliorated by increasing the colonization of <i>L. reuteri</i>
Abhisingha M ²⁸	Laboratory study	2018	-	Fifty-eight isolates consisting of <i>Lactobacillus reuteri, L. salivarius, L. mucosae, L. johnsonii,</i> and <i>L. crispatus</i> were selected and further analysed for probiotic properties, including production of antimicrobial substances and acid and bile tolerance.
Ang L. Y ²⁹	Laboratory study	2016	-	<i>L. reuteri</i> Protectis displays a significant dose-dependent antiviral activity against Coxsackievirus type A (CA) strain 6 (CA6), CA16 and EV71, but not against Coxsackievirus type B strain 2.
				CONTINUED

IABL	E 1 (CUNTINUED). S "LACTOBACILLUS	UMMARY OF THE REUTERI (DSM 179	MUSI KELE 938)", "LACI	ANI BIBLIOGKAPHY CONSIDEKED FOK INIS KEVIEW (KEYWOKDS USED: OSE INTOLERANCE", "PROBIOTICS", "MICROBIOTA", "LACTASE").
First Author	Type	 -•Year of publication 	No. of patients	Main Findings
orgensen M ³³	Laboratory study	2017	~	<i>L. reuteri</i> exhibited antifungal properties against five of the six most common oral Candida species. Further, the results reconfirms that the probiotic capacity of <i>L. reuteri</i> is strain specific.
Greifovà G ³⁴	Laboratory study	2017	-	All studied <i>L. reuteri</i> strains showed the ability to produce lactic acid, acetic acid, ethanol, reuterin and phenyllactic acid-a, a potent antifungal compound.
rhomas CM ³⁶	Laboratory study	2012	-	The identification of bacterial bioactive metabolites and their corresponding mechanisms of action with respect to immunomodulation may lead to improved anti-inflammatory strategies for chronic immune-mediated diseases.
3ene K ⁴²	Laboratory study	2017	~	Selected commensal bacterial strains are able to drive strong effector immune responses by moDCs, while in the presence of ATRA, they support the development of both tolerogenic and inflammatory moDC in a RARα-dependent manner.
ones ML ⁴⁶	Randomized controlled trial	2012	127	The deconjugation of intraluminal bile acids results in reduced absorption of non-cholesterol sterols and indicate that <i>L. reuteri</i> NCIMB 30242 capsules may be useful as an adjunctive therapy for treating hypercholesterolemia.

LI treatment. The insufficient survival and viability of L. acidophilus in commercial food products has been associated to this lack of effect by researchers¹³. *Lactobacillus rhamno-sus*, a Gram-positive lactic acid bacterium that is present in the gut microflora, seems to improve symptoms in LI subjects with a decrease in duration and frequency of diarrhea. There is limited evidence on the role of *Lactobacillus reuteri*, a Gram-positive probiotic strain that is naturally found in the gastrointestinal tracts. In a comparative study by Ojetti et al¹⁴ (2010), *L. reuteri* and tilactase, i.e., a beta-D-galactosidase obtained from Aspergillus oryzae, were proven to ameliorate gastrointestinal symptoms after lactose intake with respect to placebo. This study involved 60 subjects with lactose intolerance. However, the effects of *L. reuteri*, checked by lactose breath test were lesser than that of tilactase. Finally, although *Saccharomyces boulardii* has been extensively used as a probiotic and dietary supplement, there has been a lack of research in proving the efficacy of this yeast in the context of lactose intolerance.

Lactobacillus spp and Limosilactobacillus Reuteri

Lactobacillus spp is one of the most used probiotic families in the clinical setting and numerous food products are a source of it. In fact, the genus Lactobacillus is composed of a vast and heterogeneous range of Gram-positive, non-spore-forming, and facultative anaerobic bacteria, including *L. bulgaricus*, *L. acidophilus*, *L. rhamnosus* and *L. reuteri*^{15,16}.

Among these, one should be remembered in particular, isolated for the first time in 1962: the *L. reuteri*. This probiotic grows in atmospheres with limited presence of oxygen and is able to colonize the GI tract of humans and animals, particularly by settling in the most proximal portions of the digestive tract^{17,18}.

This ability is linked to the fact that it is able to survive bile salts and environments with acidic pH, such as the gastric one. Furthermore, *L. reuteri* is regarded as a probiotic since it meets some fundamental prerequisites for GI colonization: through adhesins it can adhere to the epithelium, ensuring host-probiotic interaction; additionally, it competes with pathogen-ic microorganisms and is a safe product to administer¹⁹⁻²¹.

The antimicrobial properties and immunomodulatory effects of *L. reuteri* strains are related to their metabolite production profile. In particular, many strains produce reuterin, a known antimicrobial compound which is a mixture of several forms of 3-hydroxypropionaldehyde (3-HPA). This molecule is spontaneously converted into acrolein, which is a cytotoxic electrophile with antimicrobial properties. Reuterin, in this way, can inhibit a series of microorganisms, in particular Gram-negative bacteria, but not only²²⁻²⁴.

Yet, as far as *L. reuteri* is concerned, the beneficial effects of this strain are expressed, not only thanks to reuterin. In fact, it has been proven to be effective against a variety of gastro-intestinal infections (such as *C. difficile, E. coli, Salmonella, H. pylori*) for the production of many other substances, including lactic acid, ethanol, acetic acid and reutericycline²⁵⁻²⁸.

L. reuteri has also shown benefits when used in viral infections with circovirus, coxsackievirus, pneumovirus, rotavirus and papillomavirus, since it secretes metabolites with antiviral components. The same is true for the antifungal properties demonstrated in the context of candidiasis²⁹⁻³³.

Some strains of *L. reuteri* are also capable of converting the amino acid L-histidine into histamine. An example derives from studies conducted on the production of histamine by *L. reuteri* 6475, which suppressed the production of tumor necrosis factor (TNF) by activated monocytes. This suppression is linked to the activation of histaminergic H2 receptors, an increase in intracellular cAMP and protein kinase A, and the simultaneous inhibition of the MEK/ERK signaling mechanism³⁴⁻³⁷.

Another property of *L. reuteri* is the ability to produce different types of vitamins, including folic acid (vitamin B9) and cobalamin (B12)^{38,39}.

The production of exopolysaccharide is another important element for the biofilm formation and the adherence of *L. reuteri* to the intestinal epithelium. This substance, in studies on porcine models, prevented the adhesion of *E. coli* to the epithelial surface and, at the same time, suppressed gene expression that would have led to the production of proinflammatory cytokines such as IL-1beta and IL-6^{40,41}. Another fundamental feature of *L. reuteri* is that of modulating the intestinal microbiota to restore tissue homeostasis in the event of a disturbed balance of the bacterial flora. To demonstrate this, a study on murine model with foxp3 mutation and consequent intestinal dysbiosis, revealed benefit in the administration of *L. reuteri* in terms of increased survival of mice and reduction of multi-organ inflammation⁴²⁻⁴⁵.

L. reuteri NCIMB 30242, for example, is able to activate the hydrolase of bile salts and the consequent increase in circulating bile acids has been proposed as an explanation for the modulation of the intestinal microbiota⁴⁶.

Furthermore, the immunomodulation of *L. reuteri* was seen to be expressed through the induction of anti-inflammatory Treg cells, but also with the suppression of Th1 and Th2 responses in mice with Treg cell deficiency^{21,45}.

Thanks to this evidence, *L. reuteri* has been used therapeutically in a variety of gastrointestinal and non-gastrointestinal conditions. An example, there are the many studies conducted on the pediatric population, where *L. reuteri* has given promising results in the context of infant colic, diarrhea, functional abdominal pain, intolerance to food, allergies, regurgitation and atopic dermatitis⁴⁷.

The main aforementioned mechanisms of action of *L. reuteri* are summarized in Figure 1.

L. Reuteri and Lactose Intolerance: What We Know So Far

On the other hand, an aspect that has not been investigated in the literature is the use of this important strain in the context of food intolerances and, in particular, lactose intolerance.

In fact, therefore, many studies have expressed themselves on the immunomodulatory, anti-inflammatory, bactericidal and microbiota regulation properties by *L. reuteri*, but only one study, conducted by Ojetti et al¹⁴, has investigated the role that this probiotic can exert in individuals with lactase deficiency.



Figure 1. This figure summarizes the different mechanisms through which *L. reuteri* exerts its beneficial properties on the gut: it is able to restore lipopolysaccharide-induced (LPS-induced) intestinal damage; its secreted factors lead to reduced release of interleukin-1a (IL-1a), interleukin-8 (IL-8) and tumor necrosis factor (TNF); particularly, reuterin (one of the most important factors secreted by *L. reuteri*) has an anti-pathogen effect towards Gram + and Gram- bacteria, fungi and yeasts.

A few years ago, prospectively studied the effect of supplementation with tilactase compared to *L. reuteri* and placebo on hydrogen breath excretion and gastrointestinal symptoms in lactose intolerant patients during a H2-LBT standard.

They enrolled 60 patients with a randomization into three groups: Tilactase group, *L. reuteri* group and Placebo group. Primary outcomes were H2-LBT normalization rate and treatment effects on mean maximum H2 concentration and gastrointestinal symptoms.

80% of Tilactase patients, 35% of *L. reuteri* patients and 0% of Placebo patients were respectively LBT-control negative.

After treatment, a significant reduction in mean peak H2 excretion was observed in the Tilactase group and in the *L. reuteri* group, compared to baseline values, while no changes were found in the Placebo group.

Significant improvement in gastrointestinal symptoms was seen in both the tilactase and *L. reuteri* groups, while no changes occurred in the placebo group.

The authors suggested that the benefit given by the probiotic was linked to the adhesion to the biofilm in proximal portions of the small intestine, where it exerted the beta-galactosidase effect.

It is important to emphasize that the administration of tilactase was, however, more effective than *L. reuteri*, and the authors attributed this to the strong interindividual variability in the level of enzymatic lactase deficiency. This led to the hypothesis that those who had severe lactase insufficiency could only benefit from supplementation with tilactase at the time of the meal, while the probiotic found space and rationality where the deficiency was milder.

It is also interesting to observe the advantages that the use of *L. reuteri* has compared to tilactase: the dosage of the exogenous enzyme must always be calibrated based on the amount of lactose contained in the food that will be eaten and must always be taken in conjunction with the meal, while the probiotic does not require dosage changes, it must be taken regardless of the amount of lactose that will be ingested and the effect of beta-galactosidase lasts until the intestinal colonization by the probiotic takes place, therefore, even once its administration is stopped.

However, no studies have yet been conducted specifying the optimal dosage and recommended duration of treatment for patients with lactose intolerance, and for a probiotic that has already proven effective in numerous contexts, its potential is worth being investigated more also in this context with future studies.

CONCLUSIONS

Our review has highlighted the potential role of Lactobacillus species, particularly *L. reuteri*, in the therapeutical management of lactose intolerance patients.

Although literature is scant on this matter, *Lactobacillus Reuteri* seems to meet the requirements to be safely used in managing symptoms of lactose intolerance.

However, no studies have yet been conducted specifying the optimal dosage and recommended duration of treatment for patients with lactose intolerance, and for a probiotic that has already proven effective in numerous contexts, its potential is worth being investigated more also in this context with future studies.

Conflict of Interest

The authors declare no conflict of interest.

Authors' Contribution

M. Brigida: manuscript writing, editing; A. Saviano: bibliographic selection; G. De Carlo: manuscript writing, editing; A. Piccioni: editing, final revision; V. Ojetti: ideation; final revision.

REFERENCES

1. Swallow DM. Genetics of lactase persistence and lactose intolerance. Annu Rev Genet 2003; 37: 197-219.

2. Seppo L, Tuure T, Korpela R, Järvelä I, Rasinperä H, Sahi T. Can primary hypolactasia manifest itself after the age of 20 years? A two-decade follow-up study. Scand J Gastroenterol 2008; 43: 1082-1087.

- 3. Di Stefano M, Veneto G, Malservisi S, Strocchi A, Corazza GR. Lactose malabsorption and intolerance in the elderly. Scand J Gastroenterol 2001; 36: 1274-1278.
- 4. Bodé S, Gudmand-Høyer E. Incidence and clinical significance of lactose malabsorption in adult coeliac disease. Scand J Gastroenterol 1988; 23: 484-488.
- 5. Pironi L, Callegari C, Cornia GL, Lami F, Miglioli M, Barbara L. Lactose malabsorption in adult patients with Crohn's disease. Am J Gastroenterol 1988; 83: 1267-1271.
- 6. Welsh JD, Griffiths WJ. Breath hydrogen test after oral lactose in postgastrectomy patients. Am J Clin Nutr 1980; 33: 2324-2327.
- 7. Pohl D, Savarino E, Hersberger M, Behlis Z, Stutz B, Goetze O, Eckardstein AV, Fried M, Tutuian R. Excellent agreement between genetic and hydrogen breath tests for lactase deficiency and the role of extended symptom assessment. Br J Nutr 2010; 104: 900-907.
- 8. Labayen I, Forga L, González A, Lenoir-Wijnkoop I, Nutr R, Martínez JA. Relationship between lactose digestion, gastrointestinal transit time and symptoms in lactose malabsorbers after dairy consumption. Aliment Pharmacol Ther 2001; 15: 543-549.
- 9. Marteau P, Flourie B, Pochart P, Chastang C, Desjeux JF, Rambaud JC. Effect of the microbial lactase (EC 3.2.1.23) activity in yoghurt on the intestinal absorption of lactose: an in vivo study in lactase-deficient humans. Br J Nutr 1990; 64: 71-79.
- 10. Marteau P, Vesa T, Rambaud JC. Lactose maldigestion. In Probiotics 1997; 65-88.
- 11. Oak SJ, Jha R. The effects of probiotics in lactose intolerance: A systematic review. Crit Rev Food Sci Nutr 2019; 59: 1675-1683.
- Marteau P, Cuillerier E, Meance S, Gerhardt MF, Myara A, Bouvier M, Bouley C, Tondu F, Bommelaer G, Grimaud JC. Bifidobacterium animalis strain DN-173 010 shortens the colonic transit time in healthy women: a double-blind, randomized, controlled study. Aliment Pharmacol Ther 2002; 16: 587-593.
- 13. Yesovitch R, Cohen A, Szilagyi A. Failure to improve parameters of lactose maldigestion using the multiprobiotic product VSL3 in lactose maldigesters: a pilot study. Can J Gastroenterol 2004; 18: 83-86.
- 14. Ojetti V, Gigante G, Gabrielli M, Ainora ME, Mannocci A, Lauritano EC, Gasbarrini G, Gasbarrini A. The effect of oral supplementation with Lactobacillus reuteri or tilactase in lactose intolerant patients: randomized trial. Eur Rev Med Pharmacol Sci 2010; 14: 163-170.
- 15. Giraffa G, Chanishvili N, Widyastuti Y. Importance of lactobacilli in food and feed biotechnology. Res Microbiol 2010; 161: 480-487.
- 16. Duar RM, Lin XB, Zheng J, Martino ME, Grenier T, Pérez-Muñoz ME, Leulier F, Gänzle M, Walter J. Lifestyles in transition: evolution and natural history of the genus Lactobacillus. FEMS Microbiol Rev 2017; 41: S27-S48.
- 17. Kandler O, Stetter KO, Kohl R. Lactobacillus reuteri sp. nov., a new species of heterofermentative lactobacilli. Zentralbl Bakteriol Hyg I Abt Orig 1980; 1: 264-269.
- 18. Frese SA, Mackenzie, DA, Peterson DA, Schmaltz R, Fangman T, Zhou Y. Molecular characterization of host-specific biofilm formation in a vertebrate gut symbiont. PLoS Genet 2013; 9: e1004057.
- 19. Valeur N, Engel P, Carbajal N, Connolly E, Ladefoged K. Colonization and immunomodulation by Lactobacillus reuteri ATCC 55730 in the human gastrointestinal tract. Appl Environ Microbiol 2004; 70: 1176-1181.
- 20. Jacobsen CN, Rosenfeldt Nielsen V, Hayford AE, Moller PL, Michaelsen KF, Paerregaard A. Screening of probiotic activities of forty- seven strains of Lactobacillus spp. by in vitro techniques and evaluation of the colonization ability of five selected strains in humans. Appl Environ Microbiol 1999; 65: 4949-4956.
- 21. Mu Q, Tavella VJ, Luo XM. Role of Lactobacillus reuteri in Human Health and Diseases. Front Microbiol. 2018; 9: 757.
- 22. Talarico TL, Dobrogosz WJ. Purification and characterization of glycerol dehydratase from Lactobacillus reuteri. Appl Environ Microbiol 1990; 56: 1195-1197.
- 23. Chen G, Chen JA. Novel cell modification method used in biotransformation of glycerol to 3-HPA by Lactobacillus reuteri. Appl Microbiol Biotechnol 2013; 97: 4325-4332.
- 24. Engels C, Schwab C, Zhang J, Stevens MJ, Bieri C, Ebert MO, McNeill K, Sturla SJ, Lacroix C. Acrolein contributes strongly to antimicrobial and heterocyclic amine transformation activities of reuterin. Sci Rep 2016; 6: 36246.
- 25. Genis S, Sanchez-Chardi A, Bach A, Fabregas F, Aris A. A combination of lactic acid bacteria regulates Escherichia coli infection and inflammation of the bovine endometrium. J Dairy Sci 2017; 100: 479-492.
- 26. Cherian PT, Wu X, Yang L, Scarborough JS, Singh AP, Alam ZA. Gastrointestinal localization of metronidazole by a lactobacilli-inspired tetramic acid motif improves treatment outcomes in the hamster model of Clostridium difficile infection. J Antimicrob Chemother 2015; 70: 3061-3069.
- 27. Ojetti V, Bruno G, Ainora ME, Gigante G, Rizzo G, Roccarina D, Gasbarrini A. Impact of Lactobacillus reuteri supplementation on anti-Helicobacter pylori levofloxacin-based second-line therapy. Gastroenterol Res Pract 2012; 2012: 740381.
- 28. Abhisingha M, Dumnil J, Pitaksutheepong C. Selection of potential probiotic Lactobacillus with inhibitory activity against Salmonella and fecal coliform bacteria. Probiotics Antimicrob Proteins 2018; 10: 218-227.
- 29. Ang LY, Too HK, Tan EL, Chow TK, Shek PC, Tham E, Alonso S. Antiviral activity of Lactobacillus reuteri Protectis against Coxsackievirus A and Enterovirus 71 infection in human skeletal muscle and colon cell lines. Virol J 2016; 13: 111.
- 30. Preidis GA, Saulnier DM, Blutt SE, Mistretta TA, Riehle KP, Major AM, Venable SF, Barrish JP, Finegold MJ, Petrosino JF, Guerrant RL, Conner ME, Versalovic J. Host response to probiotics determined by nutritional status of rotavirus-infected neonatal mice. J Pediatr Gastroenterol Nutr 2012; 55: 299-307.
- Brenner TA, Rice TA, Anderson ED, Percopo CM, Rosenberg HF. Immortalized MH-S cells lack defining features of primary alveolar macrophages and do not support mouse pneumovirus replication. Immunol Lett 2016; 172: 106-112.

- 32. Karaffova V, Csank T, Mudronova D, Kiraly J, Revajova V, Gancarcikova S, Nemcovà R, Pistl J, Vilcek S, Levkut M. Influence of Lactobacillus reuteri L26 Biocenol on immune response against porcine circovirus type 2 infection in germ-free mice. Benef Microbes 2017; 8: 367-378.
- 33. Jørgensen MR, Kragelund C, Jensen PØ, Keller MK, Twetman S. Probiotic Lactobacillus reuteri has antifungal effects on oral Candida species in vitro. J Oral Microbiol 2017; 9: 1274582.
- Greifova G, Majekova H, Greif G, Body P, Greifova M, Dubnickova M. Analysis of antimicrobial and immunomodulatory substances produced by heterofermentative Lactobacillus reuteri. Folia Microbiol 2017; 62: 515-524.
- 35. Diaz M, Ladero V, Del Rio B, Redruello B, Fernande, M, Martin MC. Biofilm-forming capacity in biogenic amine-producing bacteria isolated from dairy products. Front Microbiol 2016; 7: 591.
- 36. Thomas CM, Hong T, van Pijkeren JP, Hemarajata P, Trinh DV, Hu W. Histamine derived from probiotic Lactobacillus reuteri suppresses TNF via modulation of PKA and ERK signaling. PLoS One 2012; 7: e31951.
- 37. Rossi F, Gardini F, Rizzotti L, La Gioia F, Tabanelli G, Torriani S. Quantitative analysis of histidine decarboxylase gene (hdcA) transcription and histamine production by Streptococcus thermophilus PRI60 under conditions relevant to cheese making. Appl Environ Microbiol 2011; 77: 2817-2822.
- 38. Taranto MP, Vera JL, Hugenholtz J, De Valdez GF, Sesma F. Lactobacillus reuteri CRL1098 produces cobalamin. J Bacteriol 2003; 185: 5643-5647.
- 39. Santos F, Wegkamp A, de Vos WM, Smid EJ, Hugenholtz J. High-Level folate production in fermented foods by the B12 producer Lactobacillus reuteri JCM1112. Appl Environ Microbiol 2008; 74: 3291-3294.
- 40. Salas-Jara MJ, Ilabaca A, Vega M, Garcia A. Biofilm forming Lactobacillus: new challenges for the development of probiotics. Microorganisms 2016; 4: E35.
- 41. Chen XY, Woodward A, Zijlstra RT, Ganzle MG. Exopolysaccharides synthesized by Lactobacillus reuteri protect against enterotoxigenic Escherichia coli in piglets. (2014). Appl Environ Microbiol 2014; 80: 5752-5760.
- 42. Bene K, Varga Z, Petrov, VO, Boyko N, Rajnavolgyi E. Gut microbiota species can provoke both inflammatory and tolerogenic immune responses in human dendritic cells mediated by retinoic acid receptor alpha ligation. Front Immunol 2017; 8: 427.
- 43. Galley JD, Mackos AR, Varaljay VA, Bailey MT. Stressor exposure has prolonged effects on colonic microbial community structure in Citrobacter rodentium-challenged mice. Sci Rep 2017; 7: 45012.
- 44. Yang Y, Zhao X, Le MH, Zijlstra RT, Ganzle MG. Reutericyclin producing Lactobacillus reuteri modulates development of fecal microbiota in weanling pigs. Front Microbiol 2015; 6: 762.
- 45. He B, Hoang TK, Wang T, Ferris M, Taylor CM, Tian X, Luo M, Tran DQ, Zhou J, Tatevian N, Luo F, Molina JG, Blackburn MR, Gomez TH, Roos S, Rhoads JM, Liu Y. Resetting microbiota by Lactobacillus reuteri inhibits T reg deficiency- induced autoimmunity via adenosine A2A receptors. J Exp Med 2017; 214: 107-123
- 46. Jones ML, Martoni CJ, Prakash S. Cholesterol lowering and inhibition of sterol absorption by Lactobacillus reuteri NCIMB 30242: a randomized controlled trial. Eur J Clin Nutr 2012; 66: 1234-1241.
- 47. Lebeer S, Vanderleyden J, De Keersmaecker SC. Genes and molecules of lactobacilli supporting probiotic action. Microbiol. Mol Biol Rev 2008; 72: 728-764.