

REVIEW: GASTRIC MALIGNANCIES – MICROBES, BIOMARKERS AND MOLECULAR MECHANISMS

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Abstract – Gastric carcinoma is a very common and lethal cancer, posing a serious burden worldwide. The exact contribution of microbial risk factors for this malignancy remains to be fully understood, impairing the implementation of effective preventive strategies. In addition, insufficient knowledge of *Helicobacter pylori*-mediated gastric carcinogenesis mechanisms, absence of screening programs based on accurate biomarkers, and lack of efficient therapeutics contribute to persistent high mortality rates. This past year has witnessed the emergence of new findings on the role of microorganisms in the development of gastric cancer, on potential anti-*H. pylori* vaccines, and on novel diagnostic and prognostic biomarkers. Experimental and clinical data provided further insights into the molecular programs activated in gastric cancer, offering alternative targets for therapeutic interventions and patient stratification. This article provides an overview of the research highlights related to these topics published from April 2022 to March 2023.

Keywords: Gastric cancer, *Helicobacter pylori*, Microbiota, Epstein-Barr Virus, Cancer signaling, Cancer therapy.

INTRODUCTION

Gastric cancer is a major public health problem worldwide, with more than 1 million new cases and 768,000 deaths annually¹. Gastric cancer develops as a consequence of a long and complex interplay between microorganisms, environmental factors, and host genetic susceptibility. This heterogeneous disease presents distinct histological identities reflecting different molecular subtypes, which include tumors characterized by microsatellite instability, Epstein-Barr virus (EBV) infection, genomic stability, or chromosomal instability. Of note, molecular subtypes are associated with different clinicopathological features and prognoses. Despite advances regarding gastric cancer molecular characterization, the contribution of risk factors and mechanisms underlying gastric cancer development remain largely unexplored. Herein, we provide an overview of the latest studies addressing these issues and highlight their value for improved management of cancer patients.



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NEW METHODS FOR DETECTION OF HELICOBACTER PYLORI VIRULENCE FACTORS

Helicobacter pylori is the main risk factor for gastric cancer development. Therefore, its detection is essential for timely therapeutic intervention. Conventional methods, including PCR, urea breath test, and serological tests, are widely used for the identification of infection, however, they present limitations regarding sensitivity, specificity, and information on *H. pylori* virulence. Taking this into account, Habimana et al² developed a new method based on a modified CRISPR/Cas12a that detects *cagA* and *vacA* genes, which are important virulence markers associated with inflammation and carcinogenesis². This method can be applied directly to gastric biopsies or clinical isolates, and improves sensitivity in 16-time fold, when compared to conventional methods². A similar method was established by Liu et al³ through combination of CRISPR/Cas12a with recombinase polymerase amplification directed to the *ureB* gene³. The method showed good performance in monitoring and localizing the infection in the gastric mucosa from different clinical settings. In addition, this fast detection system requires a low amount of biological material and uses common equipment, providing an important tool for low-income countries³. With the purpose of setting up a non-invasive method, Nieuwenburg et al⁴ implemented an innovative fecal immunochemical test. Using fecal samples from 182 patients, the method proved to have a sensitivity of 94.2% and accuracy of 77.6%, which were higher than that reached by conventional stool antigen test⁴. An anti-*H. pylori* IgG ELISA was also tested as a non-invasive approach using serum of 1678 patients. This retrospective study demonstrated that in comparison with urea breath testing, the ELISA was more sensitive and less expensive, suggesting its potential in future screening programs across populations⁵.

In fact, eradication of *H. pylori* infection is an effective strategy to reduce gastric cancer incidence, as shown by Yan et al⁶ in a prospective randomized study conducted in China⁶. Upon 26.5 years of follow-up, participants receiving *H. pylori* treatment had a lower incidence of gastric cancer when compared with placebo group⁶. So far, most eradication regimens are based on the combination of antibiotics with a proton pump inhibitor. Nonetheless, antibiotic usage presents numerous challenges, namely the emergence of resistant strains and side effects on resident microbiota.

In the last year, two oral vaccines have emerged as promising approaches for *H. pylori* prevention^{7,8}. Guo et al⁷ produced a vaccine that delivers antigens of Urease, CagA, VacA and neutrophil activating proteins, using the probiotic *Lactococcus lactis*, as the delivery vector⁷. In contrast, Ghasemi et al⁸ engineered another system using *Salmonella typhimurium* as a vector for the delivery of four *H. pylori* antigens: *H. pylori* adhesion A (*HpA*), neutrophil activating protein (*Hp-NAP*), and Urease A and B⁸. Both studies^{7,8} showed that, in mice, vaccines were safe and provided protective immunity against *H. pylori* infection.

MICROBIAL FACTORS IN GASTRIC CANCER

A critical feature of *H. pylori* infection is the induction of inflammation in the gastric mucosa, which is modulated not only by bacterial virulence but also by the host immune response^{9,10}. Chronic inflammation triggered by *H. pylori* may lead to development of gastric atrophy, intestinal metaplasia, dysplasia, and cancer in a proportion of the infected individuals. By combining several techniques, including RNA-sequencing, flow cytometry, immunohistochemistry, and Chipcytometry, Koch et al¹¹ investigated the specific role of CD8+ T-cells in *H. pylori*-mediated inflammation both in mice and in humans¹¹. They showed that upon *H. pylori* infection, the T-cell population is dominated by CD8+, as an inverse correlation was detected between this cell type and bacterial colonization¹¹. Furthermore, it was demonstrated that CD8+ T-cells constitute the tissue-resident memory cells, persisting in the gastric mucosa after *H. pylori* eradication, and expanding after a secondary contact with the bacterium¹¹. Importantly, the study revealed that CD8+ T-cell infiltration was directed to the major bacterial antigen CagA, which suggests that early elimination of *cagA*-positive strains could prevent the development of premalignant lesions and reduce gastric cancer risk¹¹.

Gobert et al¹² used the INS-GAS mouse model of gastric carcinogenesis to show that *H. pylori* colonization, hyperplasia, gastrointestinal intraepithelial and immune cells were decreased

by treatment with hydroxylbenzylamine (2-HOBA). Accordingly, treatment with 2-HOBA, a scavenger of reactive aldehydes generated during inflammation, induced significant downregulation of inflammation markers, including IL-1B, TNFA, NOS2, CXCL1, IL-17 and IFN- γ , and decreased levels of DNA damage¹².

Approximately 9% of gastric cancers are positive for EBV, the great majority of them being also positive for *H. pylori*. In an attempt to assess EBV oncogenic effects, Noh et al¹³ performed a retrospective study involving 956 patients who underwent surgery for gastric cancer resection¹³. The relationship between the status of EBV and *H. pylori* with clinicopathological features was evaluated through multivariate analysis. EBV infection alone was inversely associated with overall survival, despite presenting similar clinicopathological features to those cases with coinfection¹³. Aligned with this idea, Duque et al¹⁴ investigated whether EBV infection could be associated with changes in the circulating levels of iron, facilitating *H. pylori*-induced carcinogenesis. The authors found that EBV infection is associated with higher levels of hepcidin, a negative regulator of iron entry into circulation. Data from TCGA further supported the association between EBV infection and the BMP-SMAD and the IL-1/IL-6 pathways, which control the hepcidin expression¹⁴.

Other microorganisms constituting the gastric microbiome might also influence the development of gastric cancer by increasing the amount of nitrosating compounds in the stomach. Recently, the microbial composition of paired samples from the gastric epithelium and gastric fluid was shown to change along gastric carcinogenesis, concomitantly with increased levels of nitrite and a decrease in gastric acidity¹⁵. Corroborating the existence of a cancer-associated microbiota profile, Liu et al¹⁶ identified an enrichment of opportunistic pathobionts, including *Fusobacterium*, *Parvimonas*, *Veillonella*, *Prevotella*, and *Peptostreptococcus species*, as well as a decrease of bacterial commensals as *Bifidobacterium*, *Bacillus*, and *Blautia species* in gastric cancer. Despite the application of a consistent bioinformatics pipeline to data from six distinct studies, this meta-analysis was biased regarding the geographic origin of patients, impairing more generalized conclusions¹⁶.

BIOMARKERS IN GASTRIC CANCER

Gastric cancer is frequently diagnosed at advanced stages when patients present symptoms, which limits therapeutic options and results in high morbidity and mortality. Therefore, there is an urgent need to find effective gastric cancer biomarkers for early detection of the disease. In this context, circulating proteins and metabolites emerged as potential gastric cancer biomarkers¹⁷. Deng et al¹⁷ performed a systematic review with meta-analysis to assess the value of circulating proteins, such as pepsinogen, transferrin, ferritin, and anti-*Helicobacter* IgG, as well as a panel of metabolites, including glucose, cholesterol and vitamins, α / β -carotenes, or α -tocopherol¹⁷. The analysis revealed that high levels of anti-*H. pylori* IgG, pepsinogen I (<30 μ g/L), and serum pepsinogen I/II ratio (<3) were the best candidates. As an alternative approach, Otsu et al¹⁸ screened the miRNA profile in the serum of 1206 patients, demonstrating that the combination of three miRNAs had good performance to discriminate individuals at higher risk of developing gastric cancer¹⁸.

MOLECULAR MECHANISMS IN GASTRIC CANCER

The mechanistic basis of gastric carcinogenesis has been subject of intense investigation with the purpose of identifying novel biomarkers or therapeutic targets. Among these studies, it was found that *H. pylori* infection induces expression and secretion of Laminin γ 2, an important extracellular matrix (ECM) component¹⁹. Laminin γ 2 was found to promote survival and invasion of gastric cancer cells, increasing the activity of molecules such as AKT and Src, while repressing JNK signaling. Using mutant *H. pylori* strains, clinical isolates, and gastric cancer samples, the authors further demonstrated that this effect was dependent on a functional type IV secretion system and on CagA. They propose that cells with genetic or epigenetic alterations of E-cadherin are more susceptible to *H. pylori* effects, highlighting potential benefits of eradication treatment in carriers of E-cadherin germline variants^{19,20}.

The outcome of persistent *H. pylori* infection has been associated with activation of Nuclear factor kB (NF-kB), which is a well-known transcriptional factor involved in cancer development and progression. By coupling bioinformatics analysis and *in vitro* assays, *RASAL2* was identified as a candidate gene transcriptionally regulated through NF-kB in *H. pylori*-infected tissues²¹. Consistent with this, increased *RASAL2* mRNA levels were detected in human gastric tumor tissues and mouse gastric cancer models, as well as in public data sets comprising *H. pylori* infected mice²¹. The molecular mechanism underlying oncogenic functions of *RASAL2* was found to involve the AKT/ β -catenin signaling axis, contributing to patient poor prognosis and chemoresistance²¹. *CDK1* was recently confirmed to be an additional target gene of NF-kB in response to *H. pylori* infection in gastric cancer cell lines, mouse models, and human tumor tissues²². Binding of NF-kB to the *CDK1* promoter leads to its increased expression and consequent modulation of the GSK-3 β / β -catenin pathway with impact in cell growth and apoptosis²².

Over the past decades, multiple cytokines were reported to trigger chronic gastric inflammation. High levels of IL-17A have been previously detected in serum and tumor samples of gastric cancer patients, when compared to healthy controls²³. Recently, IL-17A inhibition was found to decrease tumor growth, oxidative stress and stemness properties of cancer cells by impairing its interaction with the IL-17RC receptor and blocking the downstream NF-kB/NOX1 pathway²⁴.

Along with cytokines, oxidative stress induced by the accumulation of reactive oxygen species (ROS) has emerged as a major contributor to pro-inflammatory conditions of gastric tissues and tumors. In this regard, *H. pylori* was shown to upregulate and phosphorylate the E3 ubiquitin ligase Siah2, accelerating proteasomal degradation of the 78-kDa glucose-regulated protein (GRP78) – a powerful player in antioxidant responses. As a result, ROS accumulate in gastric cancer cells conferring proliferative advantages to that cells²⁵.

Increased levels of the cysteine-producing enzyme cystathionine γ -lyase (CTH) were also described to accompany inflammatory response to *H. pylori*, both in humans and mice²⁶. CTH enhances mitochondrial respiration and glycolysis in macrophages, supporting their metabolic activation and polarization²⁶.

The relevance of the various cell types present in the gastric tumor microenvironment was investigated by single-cell RNA-seq in a cohort of 31 gastric cancer patients with different cancer subtypes and stages²⁷. The comprehensive analysis of more than 200,000 cells, contemplating their geographic and spatial relationship, unveiled a plasma cell program involving the epithelial-resident KLF2 in diffuse-type gastric cancer. In addition, this study has identified a novel cancer-associated fibroblast subtype characterized by high INHBA-FAP expression correlated with patient poor prognosis²⁷.

Overall, the use of multidisciplinary approaches identified several molecular features, including Laminin γ 2, *RASAL2* and *CDK1*, as potential prognostic biomarkers or promising targets in gastric tumors.

MOLECULAR CLASSIFICATION AND PROGNOSTIC MARKERS IN GASTRIC CANCER

Assessing the molecular subtypes of gastric cancer may provide a crucial tool to predict clinical outcome and assist clinicians in treatment decision. To improve the knowledge on mesenchymal-subtype of gastric cancer, which encompasses microsatellite stable tumors with an epithelial-to-mesenchymal transition (EMT) phenotype, a transcriptomic survey was performed on 1000 primary tumors. Based on epigenomic profiles, Ho et al²⁸ identified a 993-gene signature able to classify these tumors, and to predict their clinical aggressiveness or resistance to targeted therapies²⁸. Notably, by analyzing DNA-regulatory elements, the authors revealed an epigenetic landscape of mesenchymal gastric cancer orchestrated by *TEAD1*²⁸. This finding pinpoints the pharmacological inhibition of *TEAD1* as a potential therapeutic solution for this gastric cancer subtype²⁸. Another study uncovered a similar value for *ARID1A*, one of the most frequently mutated genes in gastric cancer. In particular, it was verified that *ARID1A* inactivation is linked to distinct mutational signatures across molecular gastric cancer TCGA subtypes associated with NF-kB-driven proinflammatory microenvironment²⁹.

EMT signatures were investigated as prognostic biomarkers by Song et al³⁰. Based on the expression level of EMT genes, patients were stratified into two groups with distinct prognosis.

Patients harboring tumors with high levels of EMT markers had poor prognosis, while those with low levels of an EMT signature had a better prognosis and could benefit from adjuvant chemoradiotherapy³⁰.

The prognostic value of blood-derived markers, including carcinoembryonic antigen (CEA), carbohydrate antigen (CA) 19-9, alkaline phosphatase, neutrophils, hemoglobin, and lactate dehydrogenase, was also evaluated in 788 patients enrolled in a clinical trial³¹. High levels of CEA and CA 19-9 were associated with worse overall survival and event-free survival. The authors propose that monitoring these tumor markers could improve the prediction of patients' clinical outcome³¹.

Paclitaxel is a chemotherapeutic agent with anti-mitotic effects that has been used in the treatment of gastric cancer in perioperative or postoperative settings. However, there are no prognostic biomarkers to stratify patients who would benefit from paclitaxel therapy. Sundar et al³² applied a machine-learning model based on expression data of 476 genes from patients treated with Paclitaxel conjugated with the adjuvant tegafur-gimeracil-oteracil in phase III clinical trial. The trained algorithm was able to recognize resistant and sensitive patients through the assessment of a unique signature encompassing 19 genes. Upon validation in external cohorts, the authors proposed this gene signature as a predictive biomarker for paclitaxel benefit in gastric cancer³².

GASTRIC CANCER TARGETED THERAPY

Interaction between programmed cell death protein 1 (PD-1) expressed in T cells and its ligand PD-L1 expressed in tumor cells has been exploited by cancer cells to evade immune surveillance. This interplay regulates T-cell activation, and thus its blockage with specific antibodies has emerged as a potential therapy for many cancer types. Oster et al³³ investigated whether *H. pylori* infection impacted the response to immune checkpoint inhibitors. In preclinical models of colon cells, *H. pylori* infection decreased the effectiveness of cancer immunotherapy³³. This finding was further validated in a heterotopic mouse model engrafted with melanoma cells and submitted to immune checkpoint inhibitors³³. Of note, non-infected mice transplanted with fecal microbiome of infected mice retained the ability to respond to anti-PD1 and anti-CTLA4 therapy, suggesting an *H. pylori*-specific effect independent of fecal microbiota³³. The clinical relevance of *H. pylori*-mediated immunosuppression was also investigated in patients with non-small cell lung cancer (NSCLC). The analysis of *H. pylori*-seropositivity by ELISA revealed a decrease in the survival of NSCLC patients under anti-PD-1 therapy in two independent cohorts. The authors propose serologic detection of *H. pylori* as a powerful tool to predict the efficacy of immune therapy³³.

CONCLUSIONS

In the past year, important developments have emerged proposing novel approaches to reduce gastric cancer incidence and improve patient management. Highly sensitive methods were developed for *H. pylori* detection, which may be explored in large-scale screening programs using non-invasive tests. Moreover, the development of vaccines targeting *H. pylori* may become promising tools for gastric cancer prevention in the near future. Experimental and clinical evidence further elucidated the molecular effects of *H. pylori* and EBV infections, enabling the identification of new prognostic and patient stratification markers. Ultimately, this knowledge may be translated into tailored strategies for more effective gastric cancer prevention and treatment while creating new opportunities for research.

Conflict of Interest

The authors declare no conflict of interest.

Informed Consent

Not applicable.

Author's Contribution

RMF was responsible for the study concept and design. JS, IC, JF and RMF have drafted the article. JF and RMF critically reviewed the manuscript for important intellectual content. All authors approved the final version of the manuscript.

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REFERENCES

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* 2021; 71: 209-249.
2. Habimana JD, Mukama O, Chen G, Chen M, Amisshah OB, Wang L, Liu Y, Sun Y, Li AL, Deng S, Huang J, Yan XX, Rutaganda T, Mutangana D, Wu LP, Huang R, Li Z. Harnessing enhanced CRISPR/Cas12a trans-cleavage activity with extended reporters and reductants for early diagnosis of *Helicobacter pylori*, the causative agent of peptic ulcers and stomach cancer. *Biosens Bioelectron* 2023; 222: 114939.
3. Liu H, Wang J, Hu X, Tang X, Zhang C. A rapid and high-throughput *Helicobacter pylori* RPA-CRISPR/Cas12a-based nucleic acid detection system. *Clin Chim Acta* 2023; 540: 117201.
4. Nieuwenburg SAV, Mommersteeg MC, Wolters LMM, van Vuuren AJ, Eler N, Peppelenbosch MP, Fuhler GM, Bruno MJ, Kuipers EJ, Spaander MCW. Accuracy of *H. pylori* fecal antigen test using fecal immunochemical test (FIT). *Gastric Cancer* 2022; 25: 375-381.
5. Yu JH, Zhao Y, Wang XF, Xu YC. Evaluation of Anti-*Helicobacter pylori* IgG Antibodies for the Detection of *Helicobacter pylori* Infection in Different Populations. *Diagnostics (Basel)* 2022; 12.
6. Yan L, Chen Y, Chen F, Tao T, Hu Z, Wang J, You J, Wong BCY, Chen J, Ye W. Effect of *Helicobacter pylori* Eradication on Gastric Cancer Prevention: Updated Report From a Randomized Controlled Trial With 26.5 Years of Follow-up. *Gastroenterology* 2022; 163: 154-162 e153.
7. Guo L, Zhang F, Wang S, Li R, Zhang L, Zhang Z, Yin R, Liu H, Liu K. Oral Immunization With a M Cell-Targeting Recombinant *L. Lactis* Vaccine LL-pSAM-FVpE Stimulate Protective Immunity Against *H. Pylori* in Mice. *Front Immunol* 2022; 13: 918160.
8. Ghasemi A, Wang S, Sahay B, Abbott JR, Curtiss R, 3rd. Protective immunity enhanced *Salmonella* vaccine vectors delivering *Helicobacter pylori* antigens reduce *H. pylori* stomach colonization in mice. *Front Immunol* 2022; 13: 1034683.
9. Ferreira RM, Machado JC, Figueiredo C. Clinical relevance of *Helicobacter pylori* *vacA* and *cagA* genotypes in gastric carcinoma. *Best Pract Res Clin Gastroenterol* 2014; 28: 1003-1015.
10. Mejias-Luque R, Gerhard M. Immune Evasion Strategies and Persistence of *Helicobacter pylori*. *Curr Top Microbiol Immunol* 2017; 400: 53-71.
11. Koch MRA, Gong R, Friedrich V, Engelsberger V, Kretschmer L, Wanisch A, Jarosch S, Ralser A, Lugen B, Quante M, Vieth M, Vasapolli R, Schulz C, Buchholz VR, Busch DH, Mejías-Luque R, Gerhard M. CagA-specific Gastric CD8+ Tissue-Resident T Cells Control *Helicobacter pylori* During the Early Infection Phase. *Gastroenterology* 2023; 164: 550-566.
12. Gobert AP, Asim M, Smith TM, Williams KJ, Barry DP, Allaman MM, McNamara KM, Hawkins CV, Delgado AG, Blanca Piazuelo M, Rathmacher JA, Wilson KT. The nutraceutical electrophile scavenger 2-hydroxybenzylamine (2-HOBA) attenuates gastric cancer development caused by *Helicobacter pylori*. *Biomed Pharmacother* 2023; 158: 114092.
13. Noh JH, Shin JY, Lee JH, Park YS, Lee IS, Kim GH, Na HK, Ahn JY, Jung KW, Kim DH, Choi KD, Song HJ, Lee GH, Jung HY. Clinical Significance of Epstein-Barr Virus and *Helicobacter pylori* Infection in Gastric Carcinoma. *Gut Liver* 2023; 17: 69-77.
14. Duque X, Mendoza E, Morán S, Suárez-Arriaga MC, Morales-Sánchez A, Fontes-Lemus JI, Domínguez-Martínez DA, Fuentes-Pananá EM. Epstein-Barr Virus Infection Is Associated with Elevated Hcpidin Levels. *Int J Mol Sci* 2023; 24: 1630.
15. He C, Peng C, Shu X, Wang H, Zhu Z, Ouyang Y, Yang X, Xie C, Hu Y, Li N, Ge Z, Zhu Y, Lu N. Convergent dysbiosis of gastric mucosa and fluid microbiome during stomach carcinogenesis. *Gastric Cancer* 2022; 25: 837-849.

16. Liu C, Ng SK, Ding Y, Lin Y, Liu W, Wong SH, Sung JJ, Yu J. Meta-analysis of mucosal microbiota reveals universal microbial signatures and dysbiosis in gastric carcinogenesis. *Oncogene* 2022; 41: 3599-3610.
17. Deng D, Zhang Y, Zhang R, Yi J, Dong J, Sha L, Yan M. Circulating Proteins and Metabolite Biomarkers in Gastric Cancer: A Systematic Review and Meta-analysis. *Arch Med Res* 2023; 54: 124-134.
18. Otsu H, Nambara S, Hu Q, Hisamatsu Y, Toshima T, Takeishi K, Yonemura Y, Masuda T, Oki E, Mimori K. Identification of serum microRNAs as potential diagnostic biomarkers for detecting precancerous lesions of gastric cancer. *Ann Gastroenterol Surg* 2023; 7: 63-70.
19. Ferreira RM, Figueiredo J, Pinto-Ribeiro I, Gullo I, Sgouras DN, Carreto L, Castro P, Santos MA, Carneiro F, Seruca R, Figueiredo C. Activation of Laminin gamma2 by *Helicobacter pylori* Promotes Invasion and Survival of Gastric Cancer Cells With E-Cadherin Defects. *The Journal of infectious diseases* 2022; 226: 2226-2237.
20. Blair VR, McLeod M, Carneiro F, Coit DG, D'Addario JL, van Dieren JM, Harris KL, Hoogerbrugge N, Oliveira C, van der Post RS, Arnold J, Benusiglio PR, Bisseling TM, Boussioutas A, Cats A, Charlton A, Schreiber KEC, Davis JL, Pietro MD, Fitzgerald RC, Ford JM, Gamet K, Gullo I, Hardwick RH, Huntsman DG, Kaurah P, Kupfer SS, Latchford A, Mansfield PF, Nakajima T, Parry S, Rossaak J, Sugimura H, Svrcek M, Tischkowitz M, Ushijima T, Yamada H, Yang HK, Claydon A, Figueiredo J, Paringatai K, Seruca R, Bougen-Zhukov N, Brew T, Busija S, Carneiro P, DeGregorio L, Fisher H, Gardner E, Godwin TD, Holm KN, Humar B, Lintott CJ, Monroe EC, Muller MD, Norero E, Nouri Y, Paredes J, Sanches JM, Schulpen E, Ribeiro AS, Sporle A, Whitworth J, Zhang L, Reeve AE, Guilford P. Hereditary diffuse gastric cancer: updated clinical practice guidelines. *Lancet Oncol* 2020; 21: e386-e397.
21. Cao L, Zhu S, Lu H, Soutto M, Bhat N, Chen Z, Peng D, Lin J, Lu J, Li P, Zheng C, Huang C, El-Rifai W. *Helicobacter pylori*-induced RASAL2 Through Activation of Nuclear Factor- κ B Promotes Gastric Tumorigenesis via β -catenin Signaling Axis. *Gastroenterology* 2022; 162: 1716-1731.e17.
22. Zhu S, Al-Mathkour M, Cao L, Khalafi S, Chen Z, Poveda J, Peng D, Lu H, Soutto M, Hu T, McDonald OG, Zaika A, El-Rifai W. CDK1 bridges NF- κ B and beta-catenin signaling in response to *H. pylori* infection in gastric tumorigenesis. *Cell Rep* 2023; 42: 112005.
23. Wu X, Zeng Z, Xu L, Yu J, Cao Q, Chen M, Sung JJ, Hu P. Increased expression of IL17A in human gastric cancer and its potential roles in gastric carcinogenesis. *Tumour Biol* 2014; 35: 5347-5356.
24. Kang JH, Park S, Rho J, Hong EJ, Cho YE, Won YS, Kwon HJ. IL-17A promotes *Helicobacter pylori*-induced gastric carcinogenesis via interactions with IL-17RC. *Gastric Cancer* 2023; 26: 82-94.
25. Dixit P, Suratkal SS, Kokate SB, Chakraborty D, Poirah I, Samal S, Rout N, Singh SP, Sarkar A, Bhattacharyya A. Siah2-GRP78 interaction regulates ROS and provides a proliferative advantage to *Helicobacter pylori*-infected gastric epithelial cancer cells. *Cell Mol Life Sci* 2022; 79: 414.
26. Latour YL, Sierra JC, Finley JL, Asim M, Barry DP, Allaman MM, Smith TM, McNamara KM, Luis PB, Schneider C, Jacobse J, Goettel JA, Calcutt MW, Rose KL, Schey KL, Milne GL, Delgado AG, Piazuolo MB, Paul BD, Snyder SH, Gobert AP, Wilson KT. Cystathionine γ -lyase exacerbates *Helicobacter pylori* immunopathogenesis by promoting macrophage metabolic remodeling and activation. *JCI Insight* 2022; 7: e155338.
27. Kumar V, Ramnarayanan K, Sundar R, Padmanabhan N, Srivastava S, Koiwa M, Yasuda T, Koh V, Huang KK, Tay ST, Ho SWT, Tan ALK, Ishimoto T, Kim G, Shabbir A, Chen Q, Zhang B, Xu S, Lam KP, Lum HYJ, Teh M, Yong WP, So JBY, Tan P. Single-Cell Atlas of Lineage States, Tumor Microenvironment, and Subtype-Specific Expression Programs in Gastric Cancer. *Cancer Discov* 2022; 12: 670-691.
28. Ho SWT, Sheng T, Xing M, Ooi WF, Xu C, Sundar R, Huang KK, Li Z, Kumar V, Ramnarayanan K, Zhu F, Srivastava S, Isa Z, Anene-Nzulu CG, Razavi-Mohseni M, Shigaki D, Ma H, Tan ALK, Ong X, Lee MH, Tay ST, Guo YA, Huang W, Li S, Beer MA, Foo RSY, Teh M, Skanderup AJ, Teh BT, Tan P. Regulatory enhancer profiling of mesenchymal-type gastric cancer reveals subtype-specific epigenomic landscapes and targetable vulnerabilities. *Gut* 2023; 72: 226-241.
29. Xu C, Huang KK, Law JH, Chua JS, Sheng T, Flores NM, Pizzi MP, Okabe A, Tan ALK, Zhu F, Kumar V, Lu X, Benitez AM, Lian BSX, Ma H, Ho SWT, Ramnarayanan K, Anene-Nzulu CG, Razavi-Mohseni M, Abdul Ghani SAB, Tay ST, Ong X, Lee MH, Guo YA, Ashktorab H, Smoot D, Li S, Skanderup AJ, Beer MA, Foo RSY, Wong JSH, Sanghvi K, Yong WP, Sundar R, Kaneda A, Prabhakar S, Mazur PK, Ajani JA, Yeoh KG, So JB, Tan P; Singapore Gastric Cancer Consortium. Comprehensive molecular phenotyping of ARID1A-deficient gastric cancer reveals pervasive epigenomic reprogramming and therapeutic opportunities. *Gut* 2023; gutjnl-2022-328332.
30. Song J, Wei R, Huo S, Gao J, Liu X. Metastasis Related Epithelial-Mesenchymal Transition Signature Predicts Prognosis and Response to Immunotherapy in Gastric Cancer. *Front Immunol* 2022; 13: 920512.
31. Slagter AE, Vollebergh MA, Caspers IA, van Sandick JW, Sikorska K, Lind P, Nordmark M, Putter H, Braak J, Meerhoeck-Klein Kranenbarg E, van de Velde CJH, Jansen EPM, Cats A, van Laarhoven HWM, van Grieken NCT, Verheij M. Prognostic value of tumor markers and ctDNA in patients with resectable gastric cancer receiving perioperative treatment: results from the CRITICS trial. *Gastric Cancer* 2022; 25: 401-410.
32. Sundar R, Barr Kumarakulasinghe N, Huak Chan Y, Yoshida K, Yoshikawa T, Miyagi Y, Rino Y, Masuda M, Guan J, Sakamoto J, Tanaka S, Tan AL, Hoppe MM, Jeyasekharan AD, Ng CCY, De Simone M, Grabsch HI, Lee J, Oshima T, Tsuburaya A, Tan P. Machine-learning model derived gene signature predictive of paclitaxel survival benefit in gastric cancer: results from the randomised phase III SAMIT trial. *Gut* 2022; 71: 676-685.
33. Oster P, Vaillant L, Riva E, McMillan B, Begka C, Truntzer C, Richard C, Leblond MM, Messaoudene M, Machremi E, Limagne E, Ghiringhelli F, Routy B, Verdeil G, Velin D. *Helicobacter pylori* infection has a detrimental impact on the efficacy of cancer immunotherapies. *Gut* 2022; 71: 457-466.